MICROBIAL OXYGENATION OF B-NORSTEROIDS WITH Beauveria bassiana*

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Microbial oxygenation of the B-nor analogue of Reichstein's substance S and of $3\beta_1/7\alpha_2/1$ -trihydroxy-B-norpregn-5-en-20-one with *Beauveria bassiana* have been studied and the structures of the products isolated were established by spectral as well as chemical means.

In some of our previous papers we dealt with syntheses of B-noranalogues of corticoids. We described¹ synthesis of the B-noranalogue of Reichstein's substance S, studied the conditions of microbial oxygenation of some B-norpregnane derivatives² and B-norandrostane derivatives³ and recently we have described a chemical synthesis of B-norcortisol⁴ in 15-step reaction sequence. The low yields of this synthesis necessitated further studies which are presented in this paper.

A possibility which remained to be proved was microbial oxygenation of the B-noranalogue of Reichstein's substance S (1). From a number of microorganisms we investigated, only Beauveria bassiana proved suitable for our purpose. Oxygenation of the B-norderivative I with this microorganism afforded four products which were separated by column chromatography on silica gel. All of them, as detected by mass spectrometry, were products of monohydroxylation. The most lipophilic triol was obtained in 15% yield and removal of the ketol side chain by sodium bismuthate oxidation⁵ led to the known³ androstane derivative IV. This product is therefore a product of 6α-hydroxylation and has structure *II*. As was detected by spectral means the mother liquors after crystallisation of triol II contained some other component identical in polarity with the triol II. After acetylation of these mother liquors this compound was separated by chromatography as the more polar component. It proved to be a monoacetate, newly introduced hydroxyl being left unesterified under normal conditions of acetylation (room temperature, 18 h). This hydroxyl is therefore in 11B-position and comparison with the authentic⁴ B-norcortisol acetate proved this presumption. This product was therefore the triol V and the acetate has structure VI. The third product afforded after oxidative removal

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of the side chain the known³ 11 α -hydroxy derivative *IX*. It is therefore the triol *VII*. The most polar product was the triol *X*. The structure was assigned on the basis of analogy with hydroxylation of the triol *XII*, where the structure of the 15 α hydroxylated derivative *XVI* was established by ¹H-NMR spectrometry. The main product of this biotransformation was the 6 α alcohol *II* and only very small part of the substance underwent 11 β hydroxylation.

We therefore carried out further experiments with triol XII in which the position 6 is not activated. Three products of monohydroxylation were detected in the reaction mixture. Acetylation of the lipophilic component yielded a diacetate leaving the newly introduced hydroxyl unaffected. This lipophilic product is therefore the ketol XIII and the acetate has structure XIV. This was proved by transformation to the known⁴ bis(methylenedioxy) derivative XV. The second product was a B-nortetrol which afforded a triacetate on acetylation. Oxidation with sodium bismuthate yielded a 17-oxoandrostene derivative with a hydroxyl in 15 α position as follows from the ¹H-NMR spectra: The protons at C₍₁₆₎ appear as doublet of doublets (1·9 and 2·81 ppm) and decoupling experiments proved the interaction with the 15 α -proton. The tetrol is therefore the 15 α -hydroxy derivative XVI, the triacetate has structure XVII. The third, most polar product was the 11 α -hydroxy derivative XIX. This structure was securely established again by oxidative removal of the side chain to the known³ 11 α -hydroxy-B-norandrostane derivative XXI.

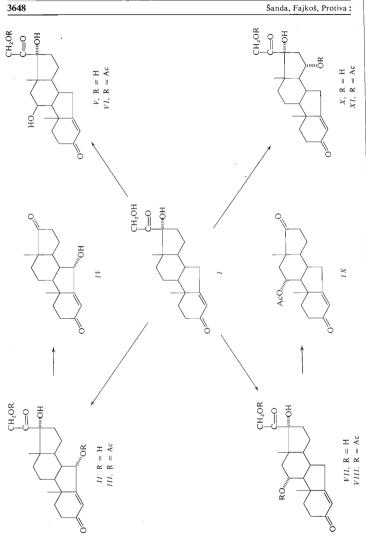
These experiments have shown that hydroxylation of the B-nortriol XII with Beauveria bassiana represents a promising pathway for synthesis of B-norcorticoids.

EXPERIMENTAL

Melting points were determined on a Kofler block. Analytical samples were dried at 80°C 0·2 Torr. Optical measurements were carried out in ethanol unless otherwise stated with an error of \pm 1°. The infrared spectra were recorded on a Zeiss UR 10 spectrometer in tetrachlormethane unless otherwise stated. The mass spectra were recorded on a AEI MS 902 mass spectrometer. The ¹H-NMR spectra were recorded on a Varian HA-100 instrument in CDCl₃-hexadeuteriodimethyl sulphoxide (1 : 1) with tetramethylsilane as internal reference at 30°C. Chemical shifts are given in ppm. The multiplicity is expressed: s (singlet), d (doublet), m (multiplet), W (width of the multiplet). Apparent coupling constants were obtained from the first order analysis. The identity of samples prepared by different routes was checked by mixture melting point determination, by thin layer chromatography (TLC) and by infrared spectra. Usual working up of an ethereal solution implies washing the solution, water, drying with magnesium sulfate and evaporation of the solvent *in vacuo*. Light petroleum refers to the fraction of b.p. 45--60°C.

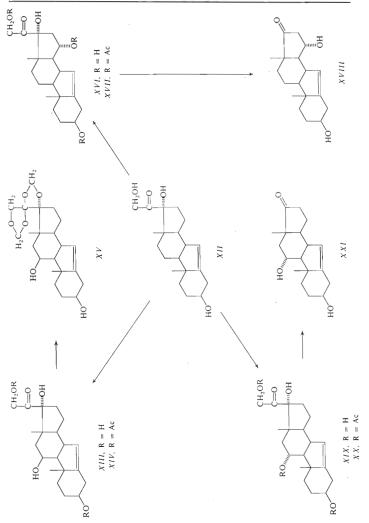
6a,17a,21-Trihydroxy-B-norpregn-4-en-3,20-dione (II)

A fermentation medium (1000 ml) containing 1% of glucose, 1.8% of corn steep oil, 0.2% of cryst. magnesium sulphate was adjusted with 10% potassium hydroxide to a pH of 6.0, sterilised at 120°C for 1 h and inoculated with 100 ml suspension of a 96 h old culture of *Beauveria bassiana*



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prepared under similar conditions from an agar culture. After 24 h at 28°C the cultivation medium was treated with a solution of the B-noranalogue of Reichstein's substance S (I) [250 mg] in dimethylformamide-ethanol (1 : 1) [5 ml]. The biotransformation was continued under the same conditions for about 30 h, when the starting steroid was no more detectable. The mycelium was then filtered off, washed with butyl acctate (3 . 500 ml) and the filtrate was extracted with the same solvent. The combined extracts were washed with a sodium carbonate solution (5%), water, dried with magnesium sulphate and the solvent was distilled off under reduced pressure. The residue was washed with light petroleum (2 . 100 ml) yielding 260 mg of the crude product of biotransformation containing, according to TLC, three components. The mixture was chromatographed on a silica gel column (50 g) in chloroform-methanol (19 : 1). Fractions with the lipophilic component were worked up to yield 35 mg of a crude product which after crystallisation from acetone gave 15 mg of the $6\alpha_17\alpha_221$ -triol *II*, m.p. 222–225°C, $[\alpha]_D^{20} + 140°$ (c 1:3). Mass spectrum: M⁺ 348. [†] H-NMR: 0·60 (s, 18-H), 1·05 (s, 19-H), 5·93 (s, 4-H), 5·08 (s, 17-OH) 4·19 and 4·59 (2 d, *J* = 19·5 Hz, 21-H), 4·44 (d, 6-H). For C₂₀H₂₈O₅ (348·4) calculated: 68·94%C, 8·10% H; found: 68·51% C, 8·11% H.

6a,17a,21-Trihydroxy-B-norpregn-4-en-3,20-dione 6,21-diacetate (111)

The triol *II* (20 mg) was acetylated with acetic anhydride (0·2 ml) in pyridine (0·4 ml) for 18 h at room temperature. The mixture was decomposed with a saturated sodium chloride solution, the product was taken into ethyl acetate and the extract was worked up. The residue was crystalised from acetone to yield 18 mg of the diacetate *III*, m.p. $167-168^{\circ}$ C. Mass spectrum: M ⁺ 432.

6α-Hydroxy-B-norandrost-4-en-3,17-dione (IV)

A solution of the dione *II* (6 mg) in 50% acetic acid (0.2 ml) was treated with sodium bismuthate (60 mg) and agitated in darkness for 3 h. The mixture was diluted with water, the product taken into ethyl acetate and the extract was worked up to yield 4 mg of the dione *IV*, m.p. 210°C, identical in all respects with the authentic³ sample.

11β,17α,21-Trihydroxy-B-norpregn-4-en-3,20-dione 21-acetate (VI)

The mother liquors obtained after crystallisation of the triol *II* from acetone (20 mg) were acetylated with acetic anhydride (0·2 ml) in pyridine (0·4 ml) overnight at room temperature. Usual working up afforded a product consisting of two components. They were separated by preparative thin layer chromatography in chloroform-methanol (19:1). The lipophilic component was the diacetate *III*, the polar compound was crystallised from acetone to yield 1·8 mg of the monoacetate *VI*, m.p. 171–173°C, identicat in all respects with the acetate prepared by acetylation of the authentic B-norcortisol *V*. Mass spectrum: M⁺ 390.

11α,17α,21-Trihydroxy-B-norpregn-4-en-3,20-dione (VII)

Elution of the chromatography after isolation of the triol *II* with the same solvent mixture (chloroform-methanol 19:1) afforded fractions with the third product of hydroxylation (30 mg). Crystallisation from acetone gave 10 mg of the B-nortriol *VII*, m.p. 205–208°C, $[a]_{10}^{20}$ +160° (c 1·2). Mass spectrum: M⁺ 348. ¹H-NMR: 0·67 (s, 18-H), 1·12 (s, 19-H), 5·74 (s, 4-H), 5·22 (s, 17-OH), 4·28 and 4·61 (2 d, J = 20 Hz, 21-H), 3·95 (m, W = 30 Hz, 11β-H). For C₂₀H₂₈O₅ (348·4) calculated: 68·94% C, 8·10% H; found: 68·61% C, 8·12% H.

11a,17a,21-Trihydroxy-B-norpregn-4-en-3,20-dione 11,21-diacetate (VIII)

The triol *VII* (20 mg) in pyridine (0·4 ml) was acetylated with acetic anhydride (0·2 ml) for 18 h at room temperature. The mixture was decomposed with a saturated sodium chloride solution, the product extracted into ethyl acetate and the solution was worked up. The residue (23 mg) after evaporation of the solvent was chromatographed over silica gel (5 g) in benzene-ether (1:1). Working up of fractions containing the acetate and crystallisation from ethyl acetate yielded 15 mg of the diacetate *VIII*, m.p. 162–163°C. Mass spectrum: M⁺ 432.

11α-Hydroxy-B-norandrost-4-en-3,17-dione 11-acetate (IX)

A solution of the diacetate VIII (9 mg) in 50% acetic acid (0.3 ml) was treated with sodium bismuthate (120 mg) and agitated in darkness for 6 h at room temperature. The mixture was diluted with ethyl acetate, treated with water and the organic layer was worked up and the residue after evaporation of the solvent (7 mg) was purified by preparative TLC in chloroform-methanol (19:1). The products were extracted with ethyl acetate to yield 4.5 mg of the starting diacetate VIII and 1.2 mg of the dione IX, m.p. $165-166^{\circ}$ C, identical in all respects with the authentic³ compound.

15α,17α,21-Trihydroxy-B-norpregn-4-en-3,20-dione (X)

Elution of the chromatography after isolation of the triols *II* and *VII* with chloroform-methanol (9 : 1) afforded fractions with the most polar product. Working up gave 30 mg of crystalls which on crystallisation from acetone yielded 10 mg of the triol *X*, m.p. $224-226^{\circ}C$, $[\alpha]_{D}^{10} + 110^{\circ}$ (c 1·1). Mass spectrum: M⁺ 348. ¹H-NMR: 0·69 (s, 18-H), 1·15 (s, 19-H), 5·66 (s, 4-H), 5·19 (s, 17-OH), 4·15 and 4·55 (2 d, J = 20 Hz, 21-H), 3·78 (m, W = 26 Hz, 15β-H). For $C_{20}H_{28}O_5$ (348-4) calculated: 68·94% C, 8·10% H; found: 68·66% C, 8·09% H.

15a,17a,21-Trihydroxy-B-norpregn-4-en-3,20-dione 15,21-diacetate (XI)

The triol X (20 mg) in pyridine (0.4 ml) was acetylated with acetic anhydride (0.2 ml) at room temperature for 18 h. The excess anhydride was removed with a saturated sodium chloride solution and the product was isolated with ethyl acetate. The solution was worked up and the residue (25 mg) was purified by preparative TLC in chloroform-methanol (19 : 1). The corresponding zone was extracted with ethyl acetate, solvent was removed and the product was regulated from acetcne to yield 15 mg of the diacetate XI, m.p. 194–195°C. Mass spectrum: M⁺ 432.

3β,11β,17α,21-Tetrahydroxy-B-norpregn-5-en-20-one (XIII)

The triol XII (500 mg) in dimethyl sulphoxide (25 ml) was submitted to hydroxylation with Beauveria bassiana as described for the diol I. The starting steroid disappeared after 24 to 30 h. The mycelium was filtered off, washed with three portions of ethyl acetate 500 ml each and the filtrate was extracted with the same solvent. The combined extracts were washed with 5% sodium hydrogen carbonate, water, dried and the solvent distilled off under reduced pressure. To remove the lipophilic components the residue was washed well with light petroleum. This crude reaction mixture (450 mg) consisted according to the TLC of three products of hydroxylation . They were separated on a silica gel column (100 g) in chloroform-methanol (19 : 1). Fractions containing the lipophilic component were worked up to yield 50 mg of a product contaminated with dimethyl sulphoxide. Repeated purification by preparative TLC in chloroform-methanol (9 : 1) and crystallisation from ethyl acetate afforded 30 mg of the tetrol XIII, m.p. $220-221^{\circ}$ C, $[\alpha]_{2}^{20}$ - 2° (α .0.96). Mass spectrum: M⁺ 350. ¹H-NMR: 0.76 (s, 18-H), 1.01 (s, 19-H), 4.18 (m, W == 8 Hz, 11α-H), 4.11 and 4.54 (2 d, J = 19 Hz, 21-H), 4.95 (s, 17-OH), 5.28 (s, 6-H), 3.32 (m, W = 30 Hz, 3α-H). For C₂₀H₃₀O₅ (350-4) calculated: 68.54% C, 8.63% H; found: 68% C, 8.61% H.

3β,11β,17α,21-Tetrahydroxy-B-norpregn-5-en-20-one 3,21-diacetate (XIV)

The tetrol XIII (3 mg) in pyridine (0.4 ml) was acetylated with acetic anhydride (0.2 ml) at room temperature for 18 h. The anhydride was decomposed with saturated sodium chloride solution and the product isolated with ethyl acetate. The residue was crystallised from acetone to yield 2 mg of the diacetate XIV, m.p. $156-157^{\circ}C$, mass spectrum: M⁺ 434.

17,20; 20,21-Bis(methylendioxy)-B-norpregn-5-en-3β,11β-diol (XV)

Paraformaldehyde (500 mg) in water (1.5 ml) and conc. hydrochloric acid (1.5 ml) was agitated at room temperature for 3 h. The tetrol XIII (40 mg) in dichloromethane (2 ml) was then added and agitated for 1 h. The mixture was treated with saturated sodium chloride solution (5 ml) and the product extracted with ethyl acetate. The extract was washed with 2% potassium carbonate with saturated sodium chloride solution, dried and the solvent removed. The residue (42 mg) was purified by preparative TLC in benzene-ether (1 : 1). The corresponding zone was extracted with chloroform to yield 19 mg of a product which on crystallisation from ethyl acetate-hexane gave 12 mg of the bis(methylendioxy) derivative XV, m.p. 230-231°C, $[\alpha]_D^{20} - 81^\circ$ (c 0.58 in chloroform). The compound was identical in all respects with the authentic compound prepared previously⁴.

3β,15α,17α,21-Tetrahydroxy-B-norpregn-5-en-20-one (XVI)

Elution of the chromatography after isolation of the tetrol XIII with chloroform-methanol (9:1) yielded fractions with the second product of hydroxylation. Working up gave 80 mg of a gumy residue containing some dimethyl sulphoxide. It was purified by preparative TLC in chloroform-methanol (9:1) and finaly chromatographed on a silica gel column (10 g) in chloroform-methanol (9:1) Working up of the corresponding fractions and crystallisation from ethyl acetate afforded 40 mg of the tetrol XVI, m.p. 238–239°C, $[zl_D^{20} - 12^\circ (c \ 1\cdot 28)$. Mass spectrum: M^+ 350. ¹H-NMR: 0·66 (s, 18-H), 0·83 (s, 19-H), 3·01 (dd, $J_{16,16} = 15$ Hz, $J_{16,15} = 9$ Hz, 16-H), 4·13 and 4·55 (2d, J = 19 Hz, 21-H), 5·59 (s, 6-H), 3·29 (m, W = 30 Hz, 3a-H), 3·88 (m, 15β-H). For C₂₀H₃₀O₅ (3504) calculated: 68·54% C, 8·63% H; found: 68·30% C, 8·52% H.

3B,15a,17a,21-Tetrahydroxy-B-norpregn-5-en-20-one 3,15,21-triacetate (XVII)

The tetrol XVI (3 mg) in pyridine (0.4 ml) was acetylated with acetic anhydride (0.2 ml) for 18 hours at room temperature. Usual working up afforded 4 mg of the triacetate XVII which resisted all attempts at crystallisation. Mass spectrum: M^+ 476.

3B,15a-Dihydroxy-B-norandrost-5-en-17-one (XVIII)

A solution of the tetrol XVI (200 mg) in 50% acetic acid (6 ml) was treated with sodium bismuthate (2.5 g) and agitated for 3 h at room temperature. The mixture was diluted with ethyl acetate and water, the upper layer was washed with hydrochloric acid and worked up. The residue was purified by preparative TLC in chloroform-methanol (9: 1). The corresponding zones were

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collected, extracted with ethyl acetate and the solvent was removed. The residue was crystallised from acetone to yield 30 mg of the diol XVIII, m.p. $252-255^{\circ}C$, $[\alpha]_{D}^{20}$ –17° (c 0.93). Mass spectrum: M⁺ 290. ¹H-NMR: 0.87 (s, 18 and 19-H), 1.90 and 2.81 (2 dd, 16-H), 4.23 (bm, 15-H), 4.54 (bs, OH), 4.88 (bd, OH), 5.65 (bs, 6-H); after addition of CD₃COOD: 0.87 (s, 18 and 19-H), 1.90 and 2.79 (2 dd, J_{16,16} = 19 Hz, J_{16,5} = 6.5 Hz + 8 Hz, 16-H), 3.34 (bs, 6-H), 4.21 (m, J_{15,14} = 9 Hz, J_{15,16} = 8 Hz + 6.5 Hz). For C₁₈H₂₆O₃ (290.4) calculated: 74.44% 9.03% H; found: 74.22% C, 9.10% H.

3B,11a,17a,21-Tetrahydroxy-B-norpregn-5-en-20-one (XIX)

Elution of the chromatography after isolation of the tetrols XIII and XVI with the same solvent mixture afforded fractions with the most polar product of hydroxylation. Working up afforded 150 mg of a solid which was purified by preparative TLC (chloroform-methanol 9 : 1) and then by column chromatography on silica gel (15 g) in chloroform-methanol (9 : 1). The corresponding fractions were worked up and the residue was crystallised from ethyl acetate to yield 100 mg of the tetrol X/X, m.p. 216–218°C. $[\alpha]_D^{20} - 58^{\circ}$ (c 1·28). Mass spectrum: M⁺ 350. ¹H-NMR: 0·57 (s, 18-H), 0·96 (s, 19-H), 4·17 and 4·58 (2 d, J = 19 Hz, 21-H), 5·04 (s, 17 α -OH), 5·25 (s, 6-H), 3·33 (m, W = 30 Hz, 3 α -H), 3·83 (m, W = 25 Hz, 11β-H). For C₂₀H₃₀O₅ (350·4) calculated: 68·54% C, 8·63% H; found: 68·22% C, 8·63% H.

3β,11α,17α,21-Tetrahydroxy-B-norpregn-5-en-20-one 3,11,21-triacetate (XX)

The tetrol XIX (3 mg) in pyridine (0.4 ml) was acetylated with acetic anhydride (0.2 ml) as described for the diacetate III. Similar working up and crystallisation from acetone gave 2 mg of the acetate XX, m.p. $191-192^{\circ}C$. Mass spectrum: M⁺ 476.

3β,11α-Dihydroxy-B-norandrost-5-en-17-one (XXI)

A solution of the tetrol XIX (200 mg) in 50% acetic acid (6 ml) was treated with sodium bismutate (2-5 g) and agitated at room temperature for 3 h. The mixture was worked up as described for the dione IV. The ethyl acetate extract was reduced to 2 ml and allowed to crystallise to yield 30 mg of the diol XXI. The mother liquors were purified by preparative TLC in chloroform-methanol (9 : 1) to yield after working up and crystallisation additional 15 mg of the diol XXI, m.p. 116 to 124° C, $[\alpha]_{D}^{20} - 80^{\circ}$ (c 1-02), identical in all respects with the authentic³ sample.

The analyses were carried out in the Analytical Laboratory of this Institute by Mr V. Štěrba, Mrs V. Rusová and Mrs E. Sýkorová under the direction of Dr J. Horáček. The infrared spectra were recorded by Mr P. Fornánek and Mrs S. Vašičková under the direction of Dr J. Smoliková. The mass spectra were recorded and interpreted by Dr L. Dolejš. The ¹H-NMR spectra were recorded and interpreted by Dr M. Buděšinský and Dr M. Synáčková. Technical assistance was provided by Mrs J. Mašková.

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